

Seawater carbonate chemistry considerations for ocean alkalinity enhancement research: theory, measurements and calculations

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Abstract. Ocean alkalinity enhancement (OAE) is a proposed marine carbon dioxide removal (mCDR) approach that has the potential for large-scale uptake of significant amounts of atmospheric carbon dioxide (CO₂). Removing anthropogenic legacy CO₂ will be required to stabilise global surface temperatures below the 1.5-2 °C Paris Agreement target of 2015. In this chapter we describe the impacts of various OAE feedstocks on seawater carbonate chemistry, as well as pitfalls that need to be avoided during sampling, storage and measurement of the four main carbonate chemistry parameters, i.e. dissolved inorganic carbon (DIC), total alkalinity (TA), pH and CO₂ fugacity (*f*CO₂). Finally, we also discuss considerations in regard to calculating carbonate chemistry speciation from two measured parameters. Key findings are 1) theoretical CO₂ uptake potential (global mean of 0.84 moles of CO₂ per mole of TA added) based on carbonate chemistry calculations are probably secondary in determining in which oceanic region OAE would be best, 2) carbonate chemistry sampling is recommended to involve gentle pressure filtration to remove calcium carbonate (CaCO₃) that might have been precipitated upon TA increase as otherwise interfering with a number of analyses, 3) samples for DIC and TA can be stabilised to avoid the risk of secondary CaCO₃ precipitation during sample storage, and 4) some OAE feedstocks require additional adjustments to carbonate chemistry speciation calculations using available programs and routines, for instance if seawater magnesium or calcium concentrations are modified.

1 Seawater carbonate chemistry revisited

25 In the following sections the basic concepts of the seawater carbonate system are laid out, together with definitions of the key parameters.

1.1 Acid-base parameters and equilibrium chemistry

The acid-base equilibrium chemistry of seawater has been described in the Guide to Best Practices for Ocean CO₂ measurements (Dickson et al. 2007), and the carbonate chemistry chapter in the Guide to Best Practices for Ocean Acidification Research and Data Reporting (Dickson 2010). These publications are open access and provide practical

guidance related to studying the carbon dioxide system in seawater. They describe tested approaches for measuring the four main parameters that are commonly used to constrain seawater acid-base chemistry and provide recommended values for the various equilibrium constants (as functions of salinity and temperature, and at 1 atm total pressure). Another key resource, although not open access, is the book ‘CO₂ in seawater: Equilibrium, Isotopes, Kinetics’ (Zeebe & Wolf-Gladrow 2001), which offers comprehensive theoretical knowledge on all aspects relevant for carbonate chemistry related research, including OAE.

The marine carbonate system can be constrained by knowing next to temperature, salinity and pressure, two carbonate chemistry parameters (Zeebe & Wolf-Gladrow 2001). Additional data such as silicic acid or phosphate concentrations might also be required, although these have typically limited influence on carbonate chemistry speciation. Four carbonate chemistry parameters are commonly measured (carbonate ion concentration measurements are less common). These are:

- DIC - the total dissolved inorganic carbon (expressed as an amount content of carbon atoms, i.e. moles per kg of seawater) in a sample of seawater (Eq. 1)
- TA - the total alkalinity (expressed as an amount content of hydrogen ions that are equivalent to the bases – defined relative to a specified equivalence point) in a sample of seawater (Eq. 2)
- pH_T - a measure of the amount content of *total* hydrogen ions in a sample of seawater (Eq. 3)
- *f*CO₂: the fugacity of carbon dioxide in air that is in solubility equilibrium with a sample of seawater (Eq. 4)

(Eq. 1) $\text{DIC} = [\text{CO}_2^*] + [\text{HCO}_3^-] + [\text{CO}_3^{2-}]$

with [CO₂*] denoting the sum of dissolved [CO₂] and [H₂CO₃].

50 (Eq. 2) $\text{TA} = [\text{HCO}_3^-] + 2[\text{CO}_3^{2-}] + [\text{B}(\text{OH})_4^-] + [\text{OH}^-] + [\text{HPO}_4^{2-}] + 2[\text{PO}_4^{3-}] + [\text{H}_3\text{SiO}_4^-] + [\text{NH}_3] + [\text{HS}^-]$
 $- [\text{H}^+]_F - [\text{HSO}_4^-] - [\text{HF}] - [\text{H}_3\text{PO}_4] + \dots - \dots$

with the last two terms denoting minor contributions of additional proton acceptors and donors.

(Eq. 3) $\text{pH}_T = [\text{H}^+]_F + [\text{HSO}_4^-]$

with [H⁺]_F describing the free hydrogen ion concentration.

55 (Eq. 4) $f\text{CO}_2 = K_0[\text{CO}_2^*]$

with K₀ denoting the temperature and salinity dependent carbon dioxide solubility, or Henry’s law constant. Another important carbonate chemistry concept is that of the calcium carbonate (CaCO₃) saturation state (Eq. 5).

(Eq. 5) $\Omega = \frac{[\text{Ca}^{2+}][\text{CO}_3^{2-}]}{K_{sp}}$

with K_{sp} describing the temperature, salinity and pressure dependent solubility product of a particular calcium carbonate morphotype such as calcite or aragonite, and [Ca²⁺] and [CO₃²⁻] being respective in-situ concentrations. When Ω is above 1 CaCO₃ precipitation is thermodynamically favoured, while below 1 it is its dissolution.

The equilibrium constants for the amount contents of the various constituents of the acid-base equilibria in a sample of seawater are essentially dependent on the major ion composition of the seawater (usually approximated by the salinity), the temperature of the seawater, and the total pressure, i.e. water depth. Hence if salinity, temperature or pressure of a seawater sample change, the various equilibrium constants will necessarily change. Thus, the pH_T and $f\text{CO}_2$ of the seawater will change as a consequence, and it is important to note the salinity, temperature and pressure at which the measurements were made. For TA and DIC this is not a problem. The individual amount contents of unionized dissolved carbon dioxide, bicarbonate ion, and carbonate ion each change as salinity, temperature and/or pressure change, but their total sum does not (the amount of carbon atoms that are in an inorganic form is conserved). Of course, this assumes that any redox reactions involving carbon do not occur on the relatively short timescales appropriate to the changes of salinity, temperature or pressure. The same applies to the sum of the individual TA components.

Any two of the five measurable parameters (DIC, TA, pH_T , $f\text{CO}_2$ and CO_3^{2-}) can – in principle – be used together with salinity, temperature, and pressure (defining suitable equilibrium constants) to calculate the state of the carbon dioxide acid-base system in a seawater sample. The parenthetical note: “in principle”, is an acknowledgement that – as a consequence of errors in the measurements of these parameters and in the measurement of the associated equilibrium constants – the results from using alternate pairs of parameters will likely not be identical (Orr *et al.*, 2018). When total alkalinity is used as a measured parameter, it implicitly incorporates all other acid-base equilibria occurring within the sample of seawater, *i.e.* in addition to the inorganic CO_2 system (Eq. 2). This is relatively straightforward if the total amount content of each acid-base system present has been measured directly, can be calculated from sample salinity, or can safely be neglected. For each acid-base system considered, the acid-dissociation constants also need to be known. A useful consequence of the TA definition is that TA does not change when CO_2 is in- or outgassing. However, there is a growing awareness that the presence of organic acids and bases in seawater can complicate the use of total alkalinity as a measurable parameter (see *e.g.*, Fong & Dickson, 2019). The contribution of such species to total alkalinity is usually neglected, especially for open ocean seawater samples.

1.2 Deffeyes diagrams

Deffeyes (1965) published a detailed description of how to use contour diagrams plotting individual seawater CO_2 system properties as a function of total alkalinity and total dissolved inorganic carbon for a seawater of a specified salinity and at a particular temperature (and pressure). Insofar as this chapter is aimed at studies of ocean alkalinity enhancement, we feel such diagrams may be useful to help visualising the compositional changes that are involved in such processes.

When an alkaline agent such as sodium hydroxide (NaOH) is used to increase total alkalinity, such addition does not change total dissolved inorganic carbon (the vertical red line, Fig. 1). Alternatively, alkalinity could be added as a soluble inorganic carbonate salt (*e.g.* ikaite, $\text{CaCO}_3 \cdot 6\text{H}_2\text{O}$, or Na_2CO_3). Both would increase TA and DIC in a ratio 2:1 (blue line, Fig. 1). As that modified water sample then equilibrates with the atmosphere, *e.g.*, takes up CO_2 from the air, DIC increases. It is

straightforward to assess the potential of 1 kg of an alkalinity-enhanced seawater to remove CO₂ from the atmosphere. It is equivalent to the change in DIC after re-equilibration to the initial *f*CO₂ prior to the alkalinity addition (horizontally moving from the tip of the alkaline agent arrow to the initial *f*CO₂). This concept takes into account that marine Carbon Dioxide Removal (mCDR) also includes scenarios in which an oceanic CO₂ source is reduced. Similar diagrams can be drawn to assess the impact of various TA additions on Ω or pH (Fig. 1b,c).

1.3 OAE impacts on seawater and potential secondary CaCO₃ precipitation

As mentioned above, ocean alkalinity can be increased by various means (see also Eisaman et al. 2023, this Guide). First, there are multiple minerals found in natural rocks which release TA upon dissolution in seawater. Dissolution kinetics depend on how finely the minerals have been milled as well as on the mineral itself (e.g. Anbeek 1992). For instance, olivine, a magnesium/iron silicate - (Mg,Fe)₂SiO₄, dissolves relatively slowly (for the purpose of OAE), on the order of weeks to months and appears to have a relatively low solubility (Monserrat et al. 2017, Flipkens et al. 2023). On the other side of the spectrum, brucite, Mg(OH)₂, dissolution rates can be on the order of hours (Moras et al. 2023a). Another way to increase oceanic TA is to add it in ‘liquid’ or aqueous form, i.e. hydroxide ions – OH⁻, which can be produced electrochemically from seawater (e.g. Rau et al. 2013, but also see Eisaman et al, 2023, this Guide).

Whichever the approach, once alkalinity is increased it raises pH, shifting carbonate chemistry speciation towards lower aqueous carbon dioxide ([CO₂]) and higher carbonate ion concentrations ([CO₃²⁻]), as well as saturation states for various calcium carbonate (CaCO₃) morphotypes such as calcite (Ω_{calc}) and aragonite (Ω_{arag}). In this context, saturation states can be considered critical thresholds, and depending on application there are levels that should not be exceeded for extended periods of times. Avoiding critical saturation thresholds is important because CaCO₃ will precipitate inorganically through a number of mechanisms at higher Ω values. Such secondary precipitation should be avoided as it reduces the oceanic uptake capacity for atmospheric CO₂ and can even lead to runaway CaCO₃ precipitation where more TA is removed than had been added, reducing typical CO₂ uptake potential by a factor of eight (Fuhr et al. 2022, Moras et al. 2022).

In seawater there are three mechanisms for inorganic CaCO₃ precipitation, 1) homogeneously - in the absence of any soluble or particulate surfaces, 2) pseudo-homogeneously – in the presence of particulate or colloidal organics, and 3) heterogeneously – in the presence of solid mineral phases (Marion et al. 2009). The critical Ω threshold beyond which CaCO₃ formation would occur is highest for homogeneous and lowest for heterogeneous precipitation, with pseudo-homogeneous in between (Morse et al. 2007). In a natural setting, homogeneous precipitation is unlikely, as seawater is hardly void of organic or inorganic particles and/or colloids. For pseudo-homogeneous CaCO₃ precipitation, the critical Ω_{arag} threshold is about 12.3 at a salinity 35 and 20°C (Marion et al. 2009). For heterogeneous precipitation, it depends on the so-called lattice compatibility between CaCO₃ and the mineral surfaces it precipitates onto. For instance, the mineral CaCO₃ has a perfect lattice compatibility, meaning that any existing pre-cursors will lead to precipitation at Ω above 1 (Zhong&Mucci

1989). The rate of precipitation is exponentially increasing with Ω , about 5-fold for a doubling in saturation state. In contrast, other mineral surfaces have lower lattice compatibilities, for instance CaCO_3 precipitation was observed for CaO and Ca(OH)_2 above an Ω of 7, while for Mg(OH)_2 rates appeared to be even further reduced (Moras et al. 2022, 2023a). Critical Ω thresholds are also influenced by grain size, salinity and dissolved organic matter concentrations (Simkiss 1964, Chave & Suess 1970, Moras et al. 2023a), and most likely temperature as well.

The risk of secondary precipitation is particularly high when TA has been increased but atmospheric CO_2 has not yet entered the ocean, i.e. Ω is still high. However, there is also the possibility to equilibrate seawater with respect to atmospheric CO_2 during TA addition which would reduce the possibility of secondary CaCO_3 precipitation as Ω would be much lower for the same amount of TA added (Bach et al. 2019). Likewise, dilution of TA-enhanced with unperturbed seawater has been shown to effectively slow/stop secondary precipitation due to a reduction in Ω (Moras et al. 2022).

1.4 TA additions and critical Ω thresholds in the surface ocean

From a practical OAE standpoint, there will always be an Ω that one will try not to go beyond for extended periods of time to minimise potential secondary CaCO_3 precipitation. How much TA can be added depends therefore on initial in-situ seawater Ω . Globally, both surface ocean Ω_{arag} and Ω_{calc} are highly correlated, at least on larger scales, with temperature and salinity. On smaller scales, there is also a biological component, i.e. photosynthesis increasing Ω and respiration decreasing it. Impacts of temperature and salinity changes on carbonate chemistry speciation are two-fold. While there is a direct effect of temperature on carbonate chemistry speciation, the bigger impact on Ω stems from the higher CO_2 solubility in low temperature waters. The resulting higher $[\text{CO}_2]$ and lower TA/DIC at atmospheric equilibrium, leads to a shift towards lower $[\text{CO}_3^{2-}]$, and hence Ω . This is because $[\text{CO}_3^{2-}]$ is approximately equal to the difference between carbonate alkalinity (CA) and DIC in the ocean (Schulz & Maher 2023). This largely explains the latitudinal gradient of surface ocean Ω (Jiang et al., 2015). Additionally, the temperature sensibility of the apparent calcium carbonate solubility product (K_{sp}') also contributes to the latitudinal Ω gradient (Mucci, 1983).

When it comes to salinity, there is also a direct effect but the major driver is that in oceanic waters DIC and TA scale with salinity (mostly driven by water evaporation and precipitation), meaning that they will be significantly reduced at lower salinities. And for a similar oceanic pH at equilibrium with atmospheric CO_2 , reducing salinity and DIC, for instance 5-fold, will reduce $[\text{CO}_3^{2-}]$ by a similar amount. In turn, this will reduce Ω , together with the salinity-related reductions in $[\text{Ca}^{2+}]$ at lower salinities. Globally, sea surface temperatures are highest in the tropics and subtropics and decrease towards higher latitudes (Figure 2A). Similarly, there is a tendency towards higher salinity at low latitudes and lower salinities at high latitudes (Figure 2B). In concert, this leads to increasing Ω from polar to tropical waters. For instance, while there are polar regions in which Ω_{arag} is close to 1 on an annual basis, in large parts of the tropical surface ocean Ω_{arag} can be close to 4

165 (Figure 2C). Finally, as mentioned earlier, there are also biological drivers of sea surface Ω , for which a good indicator is
CO₂ fugacity. Higher than current atmospheric pressures of ~ 425 μatm are indicative of respiratory CO₂ generation, reducing
 Ω , while pressures below are typically driven by photosynthetic CO₂ fixation, increasing Ω . This modulating effect of
biology on temperature- and salinity-driven global surface ocean Ω distributions can be seen by the upwelling of CO₂-
enriched deep waters in the tropical Pacific where elevated $f\text{CO}_2$ results in lower Ω_{arag} (Figure 2C, D). Altogether, this
170 determines how much TA can be added before a critical Ω threshold is reached. Assuming that Ω should not surpass a
threshold of 5 to avoid secondary CaCO₃ precipitation (Moras et al. 2022), 5 times higher TA additions would be possible at
high in comparison to low latitudes (Figure 2E).

The amount of atmospheric CO₂ that can be taken up for a certain TA increase, depends on the uptake factor η_{CO_2} ,
175 (Humphreys et al. 2018, Tyka et al. 2022). It denotes the molar ratio of the concomitant DIC increase relative to the amount
of TA added upon establishing initial seawater $f\text{CO}_2$ by air-sea gas exchange. It is mainly determined by sea surface
temperatures, with minor effects of salinity and biology. It ranges between 0.77 and 0.96, with higher values at lower
temperatures closer to the poles (Figure 2F). The resulting DIC uptake potential for global surface waters, for a critical Ω
threshold of 5, ranges between only 50 to up to 400 $\mu\text{mol kg}^{-1}$, being higher at high and lower at low latitudes (Figure 1G). It
180 is noted though, that the CO₂ uptake factor plays only a minor role, and most differences in regional CO₂ uptake potential are
driven by the amount of TA that can be added before a critical Ω threshold is reached and/or the amount of time a water
parcel stays in contact with the atmosphere (see next section). The uptake factor can be empirically estimated using available
carbonate chemistry calculation tools or calculated analytically, for which there are also dedicated routines available
(Humphreys et al. 2018)

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1.5 Cautionary note on apparent trends in regional differences for atmospheric CO₂ uptake

When looking at the distribution of global DIC uptake potential calculated for a specific OAE application (Ω threshold),
there appears to be a general trend for higher uptake at high and lower at low latitudes. However, that only applies to the
open ocean, as coastal areas are not included in the underlying GLODAP climatology. And there, the situation is likely to be
190 quite different, as these areas are heavily impacted by terrestrial influences such as freshwater input but also anthropogenic
disturbances such as nutrient runoff which affects biological processes, and hence the CO₂ uptake potential (see above), in
particular as biological activity can be orders of magnitude higher than in the open ocean. Additionally, seasonal changes to
carbonate chemistry speciation, which can be quite large, are also not captured.

195 Furthermore, it is only an uptake potential and the realised increase in DIC critically depends on regional gas transfer
velocities, i.e. how quickly atmospheric CO₂ is equilibrating with the surface ocean, and for how long the surface water with
increased TA is in contact with the atmosphere before eventually being subducted. There are regions with relatively quick
CO₂ equilibration and surface waters remaining in contact with the atmosphere on the relevant timescales, meaning that

about a year after the TA increase ~80-100% of the CO₂ uptake potential could be realised (He & Tyka 2023). In contrast,
200 there are regions with either quite slow equilibration and/or where significant portions of TA enriched surface waters lose
contact with the atmosphere for relatively long times, e.g. regions of deep water formation. Here, the realised CO₂ uptake
potential might only be 50% or less after a couple of decades (He & Tyka 2023).

Finally, a water mass that has received a TA addition might change its physical properties such as temperature, for instance
205 when moving from high to low latitudes. As such η_{CO_2} decreases along the trajectory, meaning that the CO₂ uptake potential
declines. Hence, instead of using η_{CO_2} at the site of TA addition to estimate its DIC uptake potential, it appears more
appropriate to use a global mean of 0.84.

Last but not least, it is advisable to not treat η_{CO_2} or the DIC uptake potential as a measure of OAE efficiency. The latter
210 should be assessed in two steps, i.e. first how much of a stable TA increase can be achieved, accounting for potential CaCO₃
precipitation, as well as potentially shifting natural TA source/sink terms, for instance in sediments (Bach 2023). And
secondly, how much of DIC is actually then taken up given a particular air-sea gas exchange and exposure time. Such
approach takes into account the quite different time scales of TA and DIC increases.

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2 Sampling and storage of TA-enriched water

Sampling of discrete DIC, TA, pH and $f\text{CO}_2$ should generally follow protocols described in Standard Operating Procedure
(SOP) 1 by Dickson et al. (2007). One caveat, however, is that these protocols were developed for common oceanic
carbonate chemistry conditions. Here, we discuss additional measures for sampling and sample storage that may be
220 necessary for carbonate chemistry conditions specific to OAE (i.e. high TA, high pH, and high Ω). We acknowledge that
many of the procedures described below are based on anecdotal evidence and need to be further scrutinized with increasing
maturation of OAE research and ongoing improvements of this guide.

2.1 Specific problems to consider

225 In general, there are 4 processes that can alter carbonate chemistry conditions in a sample during its collection and storage.
These are:

- 1) modifications via air-sample gas exchange (sampling)
- 2) modifications via biological activity such as respiration (storage)
- 3) modifications via the precipitation of CaCO₃ (storage)
- 230 4) modifications via diffusion of CO₂ into or out of sampling container walls (storage)

235 These four processes affect carbonate chemistry parameters differently and the effect size also depends to a large degree on the sample itself (Table 1). Sampling and storage procedures that describe how to deal with processes 1 and 2 can be found in SOP 1 by Dickson et al. (2007), such as how samples should be collected and stored, including thorough instructions of what materials are recommended to be used and how they should be prepared.

240 CaCO_3 precipitation (process 3) has received less attention in SOP 1. This is because CaCO_3 precipitation is very slow under most natural seawater carbonate chemistry conditions and is therefore not considered to affect carbonate chemistry conditions during sampling and storage of such waters. However, CaCO_3 precipitation accelerates exponentially the further Ω deviates from 1 (Zhong & Mucci 1989). Extreme Ω values which are much higher than what is commonly observed in the oceans will be frequently encountered in OAE research. Indeed, Subhas et al. (2022) experienced this problem in their OAE experiments where TA and DIC concentrations in the OAE treatment samples declined during sample storage due to CaCO_3 precipitation. The precipitation of CaCO_3 reduces TA and DIC in a 2:1 molar ratio, also changing carbonate chemistry speciation (Figure 3). As discussed above, precipitation rates critically depend on Ω but are also influenced by the presence of organic/inorganic particles as precipitation nuclei (Zhang and Mucci, 1989; Marion et al. 2009, Moras et al., 2022; Fuhr et al., 2022), the Mg concentration (Moras et al. 2023a), or the concentration of dissolved organic carbon - DOC (e.g. Chave&Suess 1970, Pan et al. 2021, Moras et al. 2023a). Since, precipitation is usually difficult to predict in terms of how long a sample might be stable, it is advisable to treat the sample in such a way that the potential problem of precipitation is minimised/mitigated.

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In the following we describe procedures to avoid changes in carbonate chemistry conditions upon sampling and storage of DIC, TA, pH, and $f\text{CO}_2$.

2.2 General sampling considerations: filtration

255 It is advisable to filter carbonate chemistry samples upon their collection to remove particles. In oceanography, the cut-off between what is considered to be dissolved or particulate has been operationally defined by passing or being retained on a GF/F filter. Such a filter has a nominal pore size of 0.7 μm , but smaller pore sizes, for instance 0.2 μm which would remove bacteria at the same time and theoretically produce a sterile sample, have also worked in the past. Filtration has multiple benefits and might even be required:

- 260 1. Particles can serve as precipitation nuclei, which catalyse CaCO_3 precipitation during storage. Removing particles via filtration will therefore help to reduce the risks of secondary precipitation.
2. Removing particles will be necessary in case CaCO_3 precipitation has already occurred in-situ to not interfere with the measurements. This is because typical analytical procedures to measure DIC and TA rely on acidifying the seawater sample during analysis, meaning that any CaCO_3 present would re-dissolve and be measured as additional
265 DIC and TA.

3. Filtration of the sample is also useful to remove substrate for biological breakdown and organisms that respire organic carbon. Respiration has limited influence on TA but strongly affects DIC, pH, $f\text{CO}_2$ and $[\text{CO}_3^{2-}]$ (Table 1). However, for samples intended to be stored longer than a few days, sterile filtration is not recommended but fixation with mercuric chloride (HgCl_2 ; Dickson et al. 2007).

270 Filtration can have negative side-effects that need to be considered and mitigated. Most obviously, filtration requires additional sampling gear which introduces contamination risks. Thus, all filtration gear must be prepared appropriately following SOP 1 in Dickson et al. (2007). The most critical issue is that filtration increases the risk of air/sample CO_2 gas exchange. This risk can be reduced when the sample is handled by gentle pressure-filtration (as opposed to vacuum filtration which is prone to gas exchange, i.e. CO_2 degassing) in a closed system. For example, the sample could be pumped via a
275 peristaltic pump using Tygon tubing (relatively impermeable for CO_2) through a membrane filter directly into the sampling bottle, filling it from bottom to top with significant overflow (Dickson et al. 2007, Schulz et al. 2017). Alternatively, the sample could be filled into a syringe (again, potential air/water gas exchange has to be minimised) and filtered through a syringe filter, again from bottom to top with overflow. CO_2 gas exchange would change DIC and hence is also a problem for pH, $f\text{CO}_2$ and $[\text{CO}_3^{2-}]$ samples, which would also change. While TA samples are not affected by air/sample CO_2 gas
280 exchange, they are sensitive to evaporation, so prolonged exposure to the atmosphere must also be avoided.

2.3 General storage considerations

Samples for DIC, TA, pH, $f\text{CO}_2$ and $[\text{CO}_3^{2-}]$ should be prepared for storage and stored as described in SOP 1 in Dickson et al. (2007). If these samples have relatively high Ω values (as a rule of thumb $\Omega_{\text{arag}} > 12$ for filtered and $\Omega_{\text{arag}} > 5$ for
285 unfiltered samples), they most likely require additional measures to avoid changes through CaCO_3 precipitation that could eventually occur during storage.

We reiterate that much of what is described below, while appearing to be logical, remains to be tested and validated in dedicated laboratory studies. Thus, care must be taken when adopting these preliminary recommendations.

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2.3.1 Preparing DIC samples for storage

DIC changes with CaCO_3 precipitation in a 1:1 molar ratio (Figure 3). Eventual CaCO_3 precipitation can be avoided by adding a strong acid (e.g. hydrochloric acid - HCl) to the sample, although precautions have to be taken (see below). HCl reduces TA and hence Ω . Such addition also decreases pH, and increases $f\text{CO}_2$ in the sample but leaves DIC concentrations
295 untouched. Increasing sample $f\text{CO}_2$ might, however, be a problem, if acid is added in excess to the added TA, $f\text{CO}_2$ will eventually rise above atmospheric levels and increase the risk of outgassing which would decrease DIC.

Hence, to mitigate CO_2 outgassing it is advisable to dose the HCl additions to just compensate the TA addition, bringing Ω down to typical seawater levels, i.e. $\Omega_{\text{arag}} \sim 2$. Also, prior to full mixing, there might be localised high $f\text{CO}_2$

300 microenvironments, hence exposure of the sample to air after acidification should be avoided, for instance by closing the lid. Again, equilibrated OAE samples most likely do not require such an acidification step (see 2.3.1).

To reduce sample dilution, it is recommended to use strong acids (e.g. HCl) at relatively high concentrations. The dilution of the DIC sample with HCl will need to be considered to calculate the DIC concentration of the undiluted sample.

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2.3.2 Preparing TA samples for storage

TA changes due to CaCO₃ precipitation occur in a 2:1 molar ratio (Fig. 3A). CaCO₃ precipitation, due to initially high Ω , can be avoided by briefly bubbling the TA sample with pure CO₂ gas. CO₂ decreases Ω and pH, and increases DIC and $f\text{CO}_2$ but
310 leaves TA unaffected (note that CO₂ is not part of the TA definition - Eq. 2). The approach was tested with 120 mL of seawater sample where high purity CO₂ gas (4.5, i.e. 99.995%) was bubbled via Tygon tubing through a clean 1 mL pipette tip into the sample for 10 seconds (Lenc et al. 2023). The CO₂ flow rate was not determined but adjusted so that the bubbling did not lead to spill over in the 125 mL sample bottle. Measured pH declined from initially ~8.7 to well below 6 within 10 seconds so that Ω_{arag} went from ~9.5 to much below 1. Under these conditions, TA loss through CaCO₃ precipitation is
315 impossible. It is noted that bringing the pH down to pH levels prior to the TA addition, i.e. around 8, would have reduced Ω_{arag} sufficiently (< 5) to avoid CaCO₃ precipitation during sample storage if the sample had been filtered.

However, there are several aspects to consider when employing this sample fixation approach. First, impurities in the CO₂ gas could contaminate the TA sample. Particle impurities can be avoided by using filters. Gas impurities (e.g. NH₃) will be
320 more difficult to remove so that it is important to use high-purity gases and test for potential TA impurities. Second, bubbling with dry CO₂ gas directly leads to evaporation of water, increasing TA. It is unclear at this stage how relevant this problem is for a short bubble burst but it could be avoided by saturating the CO₂ gas stream with H₂O before bubbling the sample as for example described in Moras et al. (2023b).

325 Finally, atmosphere-equilibrated OAE samples which have Ω levels below the critical thresholds described above, should not require CO₂ bubbling. Although it is advised to actually calculate expected sample Ω from estimates of TA and DIC prior to storage.

2.3.3 pH and $f\text{CO}_2$ samples

330 Samples for pH and $f\text{CO}_2$ cannot be stabilised with any of the methods described above. Both, the addition of HCl or CO₂ would alter the carbonate chemistry speciation, including pH and $f\text{CO}_2$. There may be methods to reduce CaCO₃ precipitation via the addition of certain inhibitors such as DOC, but such methods would need to be developed. Thus, the recommendation for pH and $f\text{CO}_2$ of samples with high Ω is to measure them immediately after collection.

335 3. Carbonate chemistry measurements of TA-enriched water

Concerning measurements of carbonate chemistry parameters, there are different needs depending on scientific research questions. Two levels of uncertainty have been proposed, one being able to detect relatively small carbonate chemistry speciation changes of ‘climate’ signals, and the other being able to monitor shorter term or spatial variability which can be considerably larger, termed ‘weather’ signals (Newton et al. 2015).

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Being able to resolve ‘climate’ signals requires measurement uncertainties equal or better than $2 \mu\text{mol kg}^{-1}$ for DIC and TA, 0.003 for pH and 0.5% for CO_2 fugacity. In comparison, for ‘weather’ signals better than $10 \mu\text{mol kg}^{-1}$ for DIC and TA, 0.02 for pH and 2.5% for $f\text{CO}_2$ are sufficient (Newton et al. 2015).

345 Concerning OAE, both thresholds could be aimed for depending on application. For example, if it is to monitor initial TA changes upon an alkalinity addition which can be on the order of several hundreds of $\mu\text{mol kg}^{-1}$ of seawater, then aiming to resolve such ‘weather’ signal with an uncertainty of better than $10 \mu\text{mol kg}^{-1}$ is probably sufficient. However, once this signal gets diluted and to monitor CO_2 ingassing and DIC increase over timescales of months an uncertainty for ‘climate’ signals must be aimed for. In fact, depending on dilution, this might even not be enough, highlighting the fact that
350 monitoring, reporting and verification (MRV), i.e. determining how many carbon credits can be assigned to a certain OAE effort in the end, is likely to require modelling (Fennel et al. 2023, Ho et al. 2023, this Guide).

In any case, a first step before measuring any carbonate chemistry sample would be to assess the accuracy (how far off are measurements with respect to certified reference materials) and precision (what is the standard deviation of replicate
355 measurements) of one’s instrumentation, allowing to estimate the overall uncertainty for each parameter in question. Another important point to consider is that OAE samples can be far off the concentrations of typical certified reference material, meaning that checking for linearity can be important (see below for details).

Finally, for any measurements described below, the starting point should be the Guide to Best Practices of ocean CO_2
360 measurements (Dickson et al. 2007). Hence, with the exception for $f\text{CO}_2$, below we are mostly referring to discrete, not in-situ, measurements, although the following recommendations should also be considered for the latter.

3.1 Measuring DIC

Acknowledging that OAE samples for DIC can be prone to air/sample gas exchange (see section 2.2.3.1), if the right
365 precautions are taken during sampling and storage, they should be straightforward to measure. This is because DIC concentrations in a ‘non-equilibrated’ OAE sample will be the same as for untreated seawater samples. However, in samples

equilibrated with the atmosphere DIC can be several hundreds of micromoles higher than typical seawater samples or certified reference material, meaning that the linearity of the measurement instrument and procedure has to be ascertained.

370 Linearity can be checked for by preparing and measuring Na_2CO_3 (ultrapure and dried at $280\text{ }^\circ\text{C}$ for 2 h) solutions of increasing DIC in Milli-Q water (ideally prepared from a concentrated stock solution), covering the required concentration range, and comparing the fractional offset of measured against theoretical concentrations for each measurement point (Figure 4). If the fraction does not change with concentration, the system response is linear and a ‘one-point calibration’ against a certified reference material will be sufficient. Using NaHCO_3 to create a DIC gradient is not ideal as it is not available in
375 ultrapure form and without further modifications it would create high $f\text{CO}_2$ samples which would need additional precautions because of potential CO_2 degassing (note that $\Delta f\text{CO}_2$ is much higher for NaHCO_3 than for Na_2CO_3 samples). Finally, it is again emphasised that sample filtration prior to measurement is most likely a crucial step to first stabilise the sample in terms of reducing the potential for CaCO_3 precipitation and second, to remove any CaCO_3 that would have precipitated prior to sampling as it would interfere with the DIC measurement (see section 2.2.2 for details).

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3.2 Measuring TA

When it comes to measuring TA, samples are likely to be enriched in comparison to the typical surface ocean. Hence, checking for linearity of the instrument setup like in the case for DIC is important. Again, this can be done by preparing suitable calibration standards, covering the required concentration range. However, unlike for relatively straight forward DIC
385 measurements this requires interpreting pH titration data on the basis of chemical acid/base equilibria in a well-defined ion matrix, called seawater (precisely defining the zero level of protons). Hence, simply using Na_2CO_3 or NaHCO_3 in Milli-Q water is not the ideal option. Instead, to cover the higher TA range, suitable amounts of NaHCO_3 could be added to seawater (Na_2CO_3 is not advisable as it is quite alkaline and might induce some sort of carbonate precipitation, impacting TA). If the TA range also requires concentrations lower than typical seawater, for instance for samples that had seen substantial amounts
390 of CaCO_3 precipitation, then this can be easily achieved by diluting natural seawater with Milli-Q water (weight by weight) and taking the change in salinity into account. Again, checking for changes in the ratio of measured to theoretical TA along the measured TA gradient will tell if the measurement system is linear, like for DIC (compare Figure 4).

A last thing to consider is that high TA samples can need considerably more titrant if the molarity of the acid is not
395 increased. Under certain circumstances this can lead to non-linearities, e.g. if the dosing unit needs calibration. Finally, as for DIC it is important that no CaCO_3 is present in the sample, hence an additional filtration step during sampling might be required.

3.3 Measuring pH

400 If pH is measured by a glass electrode, following the recommendations in Dickson et al. (2007), there should be no additional precautions required for OAE samples, whether they be of high TA and pH or high TA and DIC, hence typical seawater pH. Also, any CaCO_3 that has precipitated prior to sampling will not interfere with the measurements (potentiometric pH only). However, if CaCO_3 has precipitated post-sampling, this will have decreased the pH and an erroneous reading will be made. Concerning accuracy, potentiometric pH measurements can be less accurate than
405 spectrophotometric pH measurements (Bockmon & Dickson 2015).

For spectrophotometric measurements, one key element is the working range of the pH dye being used. It has been suggested that for sulfonephthalein indicator dyes it is between one pH unit below and above the indicator's pK_2 , (see Hudson-Heck et al. (2021) and references therein). The latter is the pH for which the concentration of the double-unprotonated form of the
410 dye is equal to that of the single-protonated one (Byrne et al. 1988). The two most commonly used pH dyes for seawater are meta-cresol purple (mCP) and thymol blue (TB), with pK_2 of ~ 8.0 and ~ 8.5 , respectively, at a temperature of 25°C and a salinity of 35, although there are a number of studies that have extended on the salinity and temperature range (see Hudson-Heck et al. (2021) and references therein). That would suggest that seawater pH ranging from 7 to 9.5 could be measured at high accuracy and precision. However, in practice accuracy can be influenced by dye impurities, and their effect can even be
415 dependent on pH. Hence, when unpurified dyes are used, it is highly recommended to check for the proper working range.

Indeed, we have found the working range to be significantly reduced for an unpurified mCP batch, for which we compared measured pH with calculated pH from measured DIC and TA (Figure 5). And while a pH deviation by 0.03 pH units at a pH of 8.5 might not seem much, it corresponds to about an offset of $30 \mu\text{mol kg}^{-1}$ in DIC or TA if this pH measurement would
420 be used for carbonate chemistry calculations without correction, which is clearly way above both the 'weather' and 'climate' thresholds.

Another option for applying corrections is by direct means without the need for ancillary TA and DIC measurements (although they are recommended to check whether corrections are actually working within the desired pH range, which is
425 actually quite important as we will see below). For that purpose, the absorbance ratios obtained with an unpurified dye need to be corrected. While the actual procedure is beyond the scope of this chapter, the reader is referred to detailed instructions in Douglas & Byrne (2017) for mCP and Hudson-Heck et al. (2021) for TB. Again, for the uncorrected pH measurements we found a considerable offset of up to 0.06 pH units (Figure 6), corresponding to a $\sim 60 \mu\text{mol kg}^{-1}$ inaccuracy in TA or DIC if this pH measurement would be used for carbonate chemistry calculations. Furthermore, the pH error is smallest around pK_2
430 and increases below and above. While the impurity correction does fix the error close to pK_2 , the observed trend remains. The fact that there is some sort of optimum curve behaviour around pK_2 points to an issue with the dye, rather than with measurements of DIC and TA. Hence, two separate linear corrections similar to the one with mCP (Figure 4) could be applied, or purified dyes being sourced.

435 Finally, as for potentiometric pH measurements the starting point for setting up spectrophotometric pH should be Dickson et al. (2007).

3.4 Measuring $f\text{CO}_2$

440 For $f\text{CO}_2$ measurements, air is being equilibrated with seawater, either via a CO_2 permeable membrane or in a so-called showerhead equilibrator. As for potentiometric pH, any CaCO_3 in suspension will not affect $f\text{CO}_2$ measurements if precipitated pre-sampling. The only difference to typical seawater measurements is that $f\text{CO}_2$ can be relatively low for ‘un-equilibrated’ OAE, e.g. below $100 \mu\text{atm}$ for a TA addition of $\sim 500 \mu\text{mol kg}^{-1}$. Hence, full equilibration of seawater with the air to be measured might take a bit longer. Other than that, we do not see any particular issues, other than the generic problems with potential CaCO_3 precipitation if samples are stored (section 2.2.3).

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4. Carbonate chemistry calculations

Carbonate chemistry calculations in an OAE context from two measured parameters can require additional considerations. One is that particular OAE applications not only change TA or eventually DIC but also the major ion composition of seawater. For instance, for calcium-based OAE, with for instance calcium oxide or hydroxide - CaO and $\text{Ca}(\text{OH})_2$, for each 450 mol of TA half a mol of Ca^{2+} will be added. This is the same for magnesium-based OAE, for instance with $\text{Mg}(\text{OH})_2$ or olivine (forsterite). For example, increasing TA by $500 \mu\text{mol kg}^{-1}$ would increase both calcium and magnesium ion concentrations by $250 \mu\text{mol kg}^{-1}$, i.e. by 2.5% and 0.5% , respectively, over seawater background levels at a salinity of 35.

For calcium-based OAE this means that calculations of calcium carbonate saturation states have to factor in the increase in 455 the calcium to salinity ratio. In other words, using standard settings in various carbonate chemistry speciation calculation programs, e.g. CO2SYS, Seacarb, PyCO2SYS (for an overview see Orr et al. 2015), which derive calcium ion concentrations from salinity, will result in wrongly calculated saturation states. Hence, they would need to be calculated in a separate step from calculated carbonate ion and actual calcium concentrations (e.g. Moras et al. 2022), the programs being adapted, or special functions used (for instance available for Seacarb).

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Furthermore, making changes to the matrix of the major ions in seawater also affects acid/base equilibria, i.e. stoichiometric dissociation constants such as for carbonic acid. Hence, one may also need to apply corrections to K_1 and K_2 for carbonic acid (Ben-Yakoov & Goldhaber 1972) which then can be used to calculate carbonate chemistry speciation, either by hand (Zeebe & Wolf-Gladrow 2001) or by adapting available programs. Furthermore, the solubility product of calcium carbonate, 465 K_{sp} , needs to be corrected which can be achieved by using the specific magnesium to calcium ratio in a seawater sample (Tyrell & Zeebe 2004). Finally, the effects of major (and minor) ion composition changes on carbonate chemistry speciation can also be assessed by Pitzer modelling (for an overview see Turner et al. 2016), and there have been several programs and

functions made available by the SCOR Working Group 145, MARCHEMSPEC: Chemical Speciation Modelling in Seawater to Meet 21st Century Needs (<http://marchemspec.org/>).

Key recommendations

Calcium carbonate (CaCO_3) will precipitate in seawater when the saturation state is beyond certain thresholds (critically depending on the presence of organic matter or mineral precursors, among others). Hence, when it comes to determine carbonate chemistry speciation for samples with significantly increased total alkalinity (TA) at regular dissolved inorganic carbon (DIC) concentrations, CaCO_3 that has precipitated prior to sampling has to be removed for most measurements (TA, DIC, spectrophotometric pH) as it interferes with the analytical procedures. Gentle pressure filtration, avoiding gas exchange of the sample with air, is recommended. Furthermore, if samples are to be stored prior to analysis, CaCO_3 precipitation needs to be prevented, which can be achieved for TA (lowering the calcium carbonate saturation state by brief sparging with CO_2), and DIC (acidifying the sample to reduce TA to natural pre-OAE conditions). Samples for $f\text{CO}_2$ and pH cannot be stabilised and should therefore not be stored. Any sample that has been stored should be checked visually for potential CaCO_3 precipitation on container walls or in suspension. For carbonate chemistry speciation calculations from two measured parameters, if experimental treatments have changed the Mg^{2+} to Ca^{2+} ratio in seawater this should be considered by applying corrections to stoichiometric equilibrium constants. Also, experimental changes to Ca^{2+} concentrations mean that readily available calculation routines will report a wrong CaCO_3 saturation state as simply related to salinity. Again, corrections have to be made.

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Code/Data availability

No original data has been produced for this manuscript and procedures to reproduce figures are described or referenced.

Author contributions

475 The manuscript was conceptually developed by all authors. Sections 1.1 and 1.2 were led by AGD, section 2 by LTB and the rest by KGS. All authors contributed to writing, graphing and editing.

Competing interests

Competing interests are declared in a summary for the entire volume at: <https://sp.copernicus.org/articles/sp-bpoe-cisummary.zip>.

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Table 1: Direction of change in a measured carbonate chemistry parameter due to various processes. Note that for process 1 the direction depends on whether the sample is over or undersaturated with respect to atmospheric CO₂. For process 4 the direction depends on whether CO₂ is diffusing from the walls into the sample or vice versa. Also, the direction of TA change in process 2 depends on which form of inorganic nutrients are being released during organic matter remineralisation

	Air-sample CO ₂ exchange (process 1)	Respiration/Remineralisation (process 2)	CaCO ₃ precipitation (process 3)	Absorption and diffusion of CO ₂ to/from walls (process 4)
TA	N/a	+ / -	-	N/a
DIC	+ / -	+	-	- / +
pH	- / +	-	-	+ / -
<i>f</i> CO ₂	+ / -	+	+	- / +
[CO ₃ ²⁻]	- / +	-	-	+ / -

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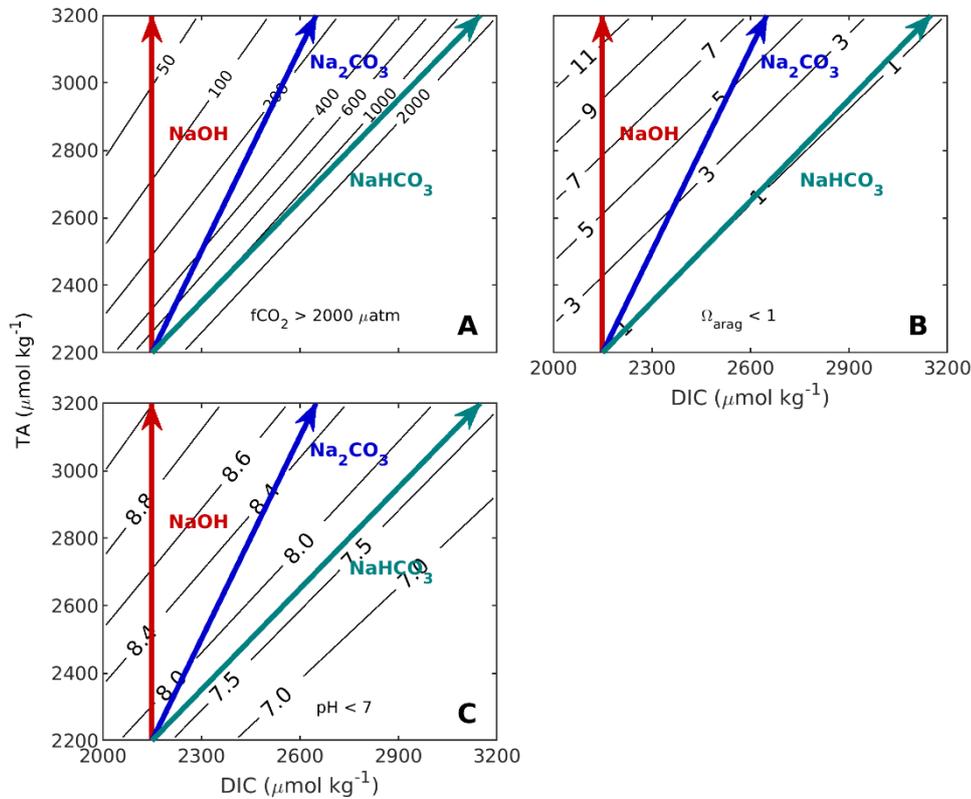


Figure 1: Implications of changes in C_T and A_T on the properties of undersaturated seawater appropriate to the California Current region. ($S = 33.5$, $t = 10$ °C, $A_T = 2200$ $\mu\text{mol kg}^{-1}$, $C_T = 2150$ $\mu\text{mol kg}^{-1}$). (A) $f(\text{CO}_2)$; (B) aragonite saturation state (Ω_{arag}); (C) pH_T . Calculations were carried out with the Matlab version of CO2SYS (Pierrot et al., 2006) using the stoichiometric dissociation constants for carbonic acid from Sulpis et al. (2020), for sulphuric acid by Dickson et al. 1990, and total boron from Uppström (1974). The red line indicates the effect of adding NaOH, the blue line of adding Na_2CO_3 and the teal line of adding NaHCO_3 . If a different initial seawater is chosen, this whole grouping (red, blue and teal lines) can be translated (moved without any distortion or rotation) so that its initial position is elsewhere on these figures.

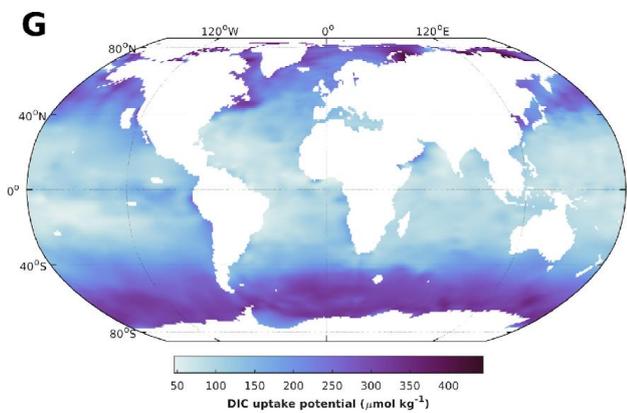
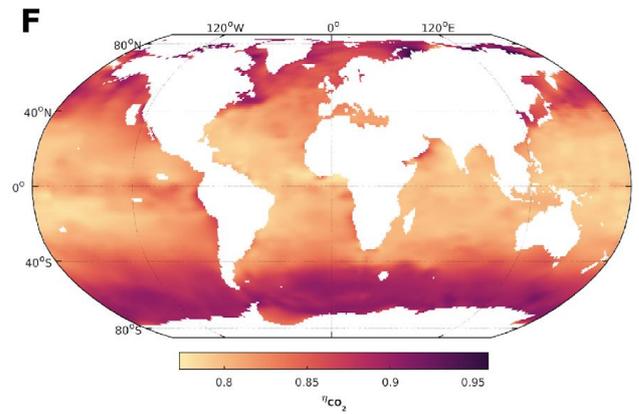
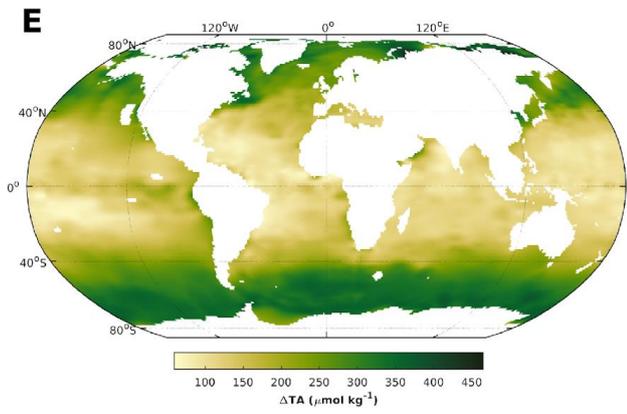
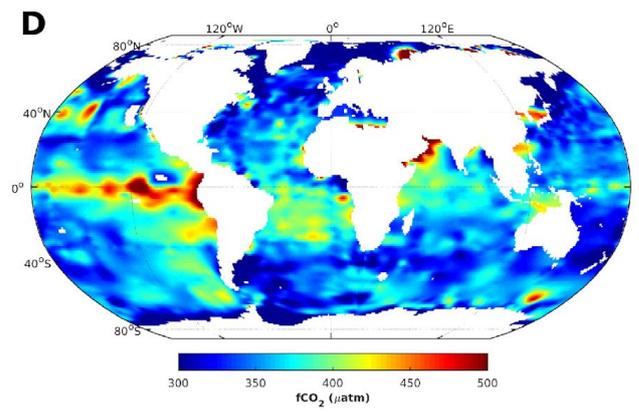
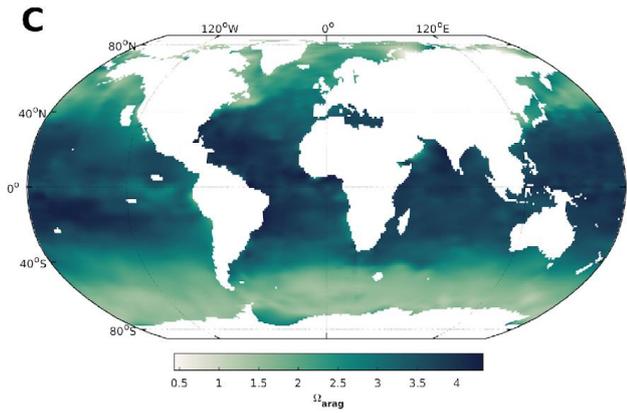
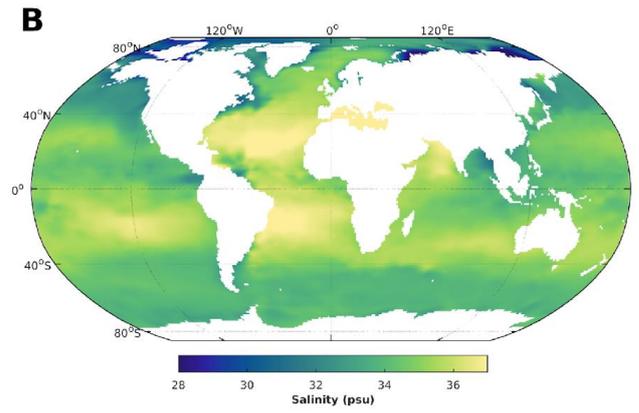
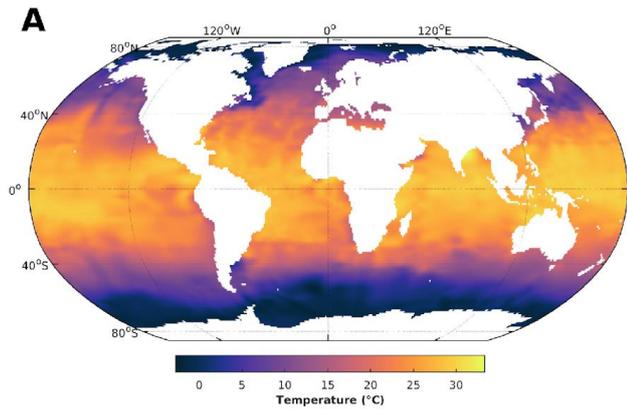
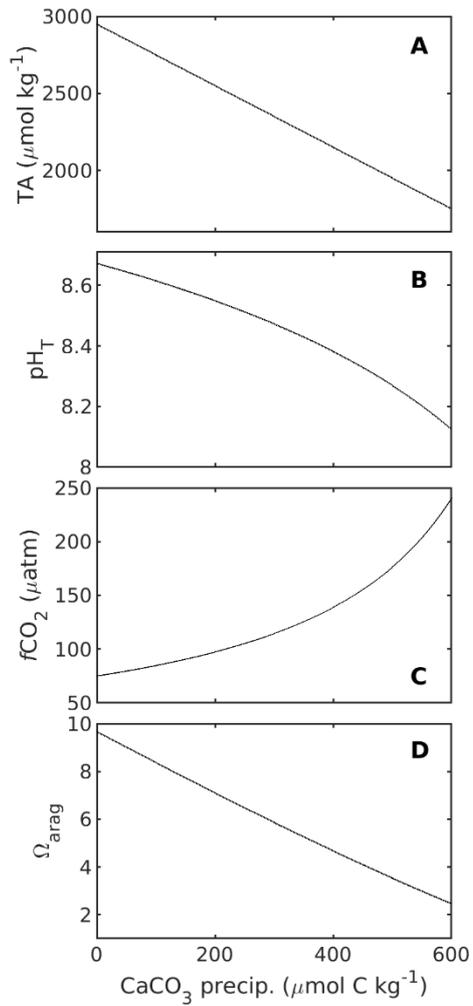


Figure 2: 1x1 degree GLODAP climatology (Lauvset et al. 2016) for surface ocean (upper 30 m) temperature (A), salinity (B), aragonite saturation state (C) and $f\text{CO}_2$ (D), the latter two calculated from GLODAP dissolved inorganic carbon and total alkalinity using the stoichiometric dissociation constants for carbonic acid from Sulpis et al. (2020), for sulphuric acid by Dickson et al. 1990, and total boron from Uppström (1974), the concentration by which total alkalinity can be increased in order to reach an aragonite saturation state of 5, ΔTA (E), the CO_2 uptake factor ηCO_2 (F), and the resulting increase in DIC by uptake of atmospheric CO_2 upon air-sea equilibration (G). For details see text.

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675 **Figure 3:** Changes in TA (A), pH_T (B), fCO₂ (C) and Ω_{arag} (D) in response to CaCO₃ precipitation, i.e. DIC removal. Carbonate chemistry calculations were for a salinity of 35 at 20 °C as described in Figure caption 1. Starting TA and DIC were 2950 and 2100 μmol kg⁻¹. Please note that for 1 mol of CaCO₃ precipitated, 1 mol of DIC is consumed.

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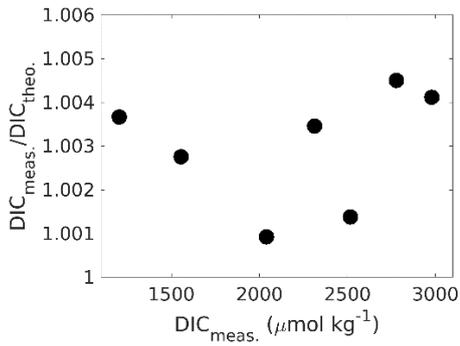


Figure 4: Measured DIC concentrations versus the ratio of measured to theoretical DIC, showing that DIC is measured between ~0.1 to 0.4% higher than theoretically predicted from self-prepared Na₂CO₃ solutions, without obvious trend across the DIC range. If the instrument response would not be linear, this ratio would change with concentration changes. When linear, the mean ratio of measured to theoretical DIC allows for correcting measured concentrations for any accuracy bias.

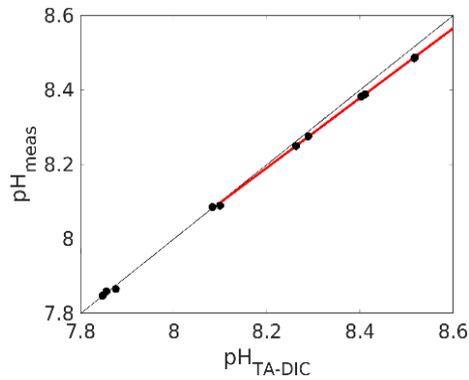
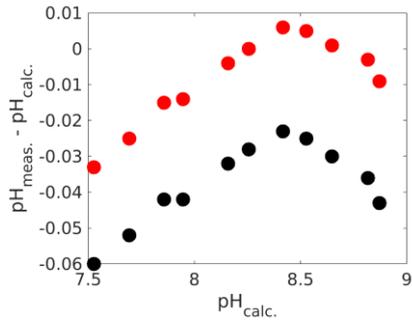


Figure 5: Calculated (from measured DIC and TA, see Figure caption 1 for details) vs. measured pH (total scale) using an unpurified batch of mCP. The red line denotes a linear fit through the data above a pH of 8.1.



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Figure 6: Calculated pH (from measured DIC and TA) versus the difference between measured and calculated pH (total scale). The black dots are values prior to the impurity correction and the red ones after (see text for details).